# **Ensembl Plants primer**

In the following paragraphs we will give a basic introduction on how to use the Ensembl Plants website to find and download data. This primer is written for the Ensembl Plants website (<u>www.plants.ensembl.org</u>) with a special focus on the wheat data section of Ensembl Plants. The basic steps described here can also be used when browsing other species.

#### **Disclaimer:**

The Ensembl Plants website is continuously being updated. We will strive to keep this document up-to-date, but if you should find that a certain aspect of the website is no longer working (or working differently) than how it is described here please consult the help pages on the Ensembl Plants website (http://plants.ensembl.org/info/website/index.html).

# a) What is on the Ensembl Plants website?

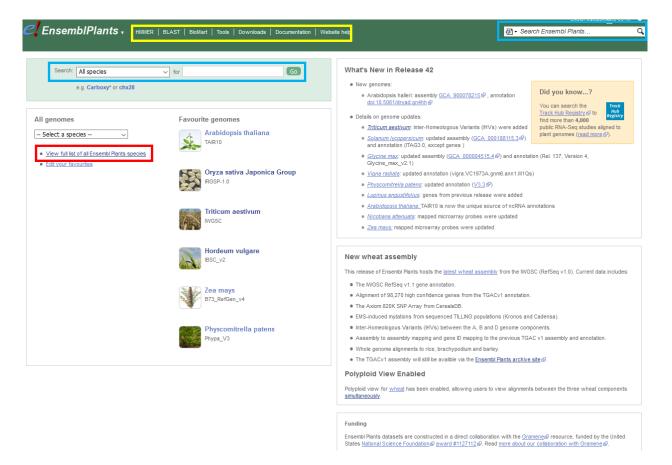
## A brief overview of available data and how to access it

The Ensembl Plants homepage (www.plants.ensembl.org) allows users to access data in different ways. There are two search bars (blue boxes in **Figure 1**) that allow the user to query either the entire database or a single species for e.g. genomic positions or genes. At the moment, the gene models for wheat (IWGSC RefSeqv1.1) are not associated with a lot of metadata, i.e. a search for the well-known vernalisation gene *Vrn1* would yield hits to *Arabidopsis thaliana* and *Brassica rapa*, but not to *Triticum aestivum*, even though the gene does exist within the database. Instead, gene names need to be queried using their designation, e.g. *TraesCS5A02G391700* for *Vrn1-A*.

The database can also be searched via HMMER (protein) or via BLAST (both DNA and protein) (yellow box in **Figure 1**). The first one uses Hidden Markov Models (HMM) to discover even remote homologues using a probabilistic method (see <a href="http://hmmer.org/">http://hmmer.org/</a> for more information), while the second one employs the BLAST algorithm (<a href="http://https://blast.ncbi.nlm.nih.gov/Blast.cgi">http://https://blast.ncbi.nlm.nih.gov/Blast.cgi</a>) to identify the most similar sequences. BLAST is currently the best option to query a gene in wheat, as the gene database for wheat is superior to the protein database, e.g. a BLAST search for *Vrn1* would lead the user to the correct wheat gene.

Users can download large quantities of specific data, e.g. all genes within a certain interval using the BioMart tool (under the "BioMart" tab, yellow box in **Figure 1**) or download raw data from an FTP server (under the "Downloads" tab, yellow box in **Figure 1**). These and other tools can be also accessed from the tools section (under the "Tools" tab, yellow box in **Figure 1**).

The documentation and help sections are well worth reading and explain the tools and data in more detail (under the "Documentation" and "Website help" tabs, yellow box in **Figure 1**). For usage tutorials go to <u>http://www.ensembl.org/info/website/tutorials/index.html</u>.



## Figure 1: Ensembl Plants homepage

This is a screenshot of the Ensembl Plants website. It contains quick links to the most used species, a search box for user queries as well as information about improvements from the current release. The link to the full list of species is highlighted with a red box.

As of November 2019, there is data for 67 species deposited on the Ensembl Plants website. This includes dicots (e.g. *Arabidopsis, Brassica*, and tomato), monocots (e.g. wheat, barley, and rice) as well as data for some algae and mosses. You can view the list of species as well as the type of data available for each by clicking the "View full list of all Ensembl Plants species" link in the middle left of the website (red box in **Figure 1**).

If you follow the link to view the full list of species on Ensembl Plants you are presented with a table listing (see **Figure 2**):

- 1. The species name
- 2. The species classification
- 3. The UniProt Taxon ID of the species
- 4. The name of the current genome assembly
- 5. A link to the genome assembly on the European Nucleotide Archive (ENA)
- 6. A list of the type of data available on Ensembl Plants for each species, e.g. variation data

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Aegliops tauschli	Liliopsida	<u>37682</u>	ASM34733v1	GCA_000347335.1			0	٢	٢	
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Arabidopsis lyrata	eudicotyledons	<u>81972</u>	v.1.0	GCA_000004255.1			٥	٢	٢	-
Arabidopsis thaliana	eudicotyledons	3702	TAIR10	<u>GCA_000001735.1</u>	0	٥	0	٥	٢	0
Brachypodium distachyon	Liliopsida	15368	v1.0	GCA_000005505.1	0	٢	0	٥	٢	-
Brassica oleracea	eudicotyledons	<u>109376</u>	v2.1	GCA_000695525.1		-	٥	٥	٢	-
Brassica rapa	eudicotyledons	<u>51351</u>	IVFCAAS/1	GCA_000309985.1	-	-	0	٢	٢	-
Chlamydomonas reinhardtil	Chlorophyta	3055	v3.1	GCA_000002595.2			٥	٥	٢	٥
Cyanidioschyzon merolae	Rhodophyta	280699	ASM9120v1	GCA_000091205.1	-	-	0	٢	٢	٢
Glycine max	eudicotyledons	<u>3847</u>	V1.0	GCA_000004515.1			ø	٢	٥	
Hordeum vulgare	Liliopsida	<u>112509</u>	ASM32608/1	GCA_000326085.1	0	٢	0	٢	٥	-
Leersia perrieri	Liliopsida	77586	Lperr_V1.4	GCA_000325765.3			ø	٢	٢	
Medicago truncatula	eudicotyledons	3880	MedtrA17_4.0	GCA_000219495.2			٥	٢	٢	

## Figure 2: Full list of all species on Ensembl Plants

This is a screenshot of the full list of species included on Ensembl Plants. There are links to the official UniProt Taxon ID and the genome assembly used on Ensembl Plants. The available data for each species is divided into six categories, e.g. variation data. A green tick mark indicates whether data of a certain type is available for a species.

After selecting your species of interest (in our case wheat; *Triticum aestivum*) you are presented with a new window containing lots of important information, e.g. details about the genome assembly (red box in **Figure 3**). This is crucial information as it helps users to understand what type of data is actually present in the database. Again, there are search bars for user queries as well.

C EnsemblPlants - HMMER   BLAST   BioMart   Tools   Downloads   Documentation	Website help	nikolai.adamsko@jic.ac.uk
Triticum aestivum (IWGSC) <del>•</del>		
Search Search Go e.g. TraceCS3002G273600 or 3D:2585940-2634711 or Carboxy* About Triticum aestivum Triticum aestivum (bread wheat) is a major global cereal grain essential to human nutrition. Wheat was one of the fit size estimated at -17 Gb, composed of three closely-related and independently maintained genomes that are the res A-genome donor) ho produce modern bread wheat. Taxonomy D 455510 Data source International Wheat Genome Sequencing Consortium(9)	archive (? ste. st cereals to be domesticated, originating in the <u>fertile crescent</u> (? It of a series of naturally occurring hybridisation events. The and	ry other plant from release 31 is available in the new <u>Ensembl Plants</u>
More information and statistics	Download genes, cDNAs, ncRNA, proteins - F	ADH PS/ Example gene
Comparative genomics What can I find? Homologues, gene trees, and whole genome alignments across multiple species. More about comparative analyses Phylogenetic overview of gene families Download alignments (EMF) Genomic alignments (E§] (Shows ] Syntenies [1] [Shows ]	Variation         What can I find? Short sequence variants.         Image: Comparison of the product of	ATCGAGCT ATCCAGCT ATCGAGAT Example variant
Regulation         What can I find? Microarray annotations.         Image: State of the sta		

## Figure 3: Species specific information

This screenshot shows species specific information for wheat. It contains details on the genome assembly, gene annotation, variation data, etc.

# b) The layout of Ensembl Plants

## The Location tab

So far, we have been navigating the main page as well as the tool and help pages of the Ensembl Plants website, which represent part of the first layer of the website.

Once we query the database we are presented with a list of hits (see **Figure 4**). For every query hit the user is presented with there are several options on how to continue. These usually are:

- 1. Gene ID; if the query was a gene or a scaffold with a gene, then this displays the ID of the gene in the database; in case of multiple genes on the same scaffold, a separate hit is shown for every gene.
- 2. Species; if querying from the main website with all species, this allows to filter the results by species.
- 3. Location; the genomic location of the query, which can be either a scaffold or a chromosome location.
- 4. Gene trees; if the query was a gene or a scaffold with a gene, then this displays the name of the gene tree created with Plant Compara or Pan-taxonomic Compara (see section c "The power of Compara").

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	Description	Heat shock protein 101 [	Source:UniProtKB/TrEMBL;Acc:Q9SPH	4]		
Share this page	Gene ID	TraesCS3D02G273600				
👫 Bookmark this page	Species	Triticum aestivum				
Ensembl Plants is produced in	Location	3D:379535906-37953982	27			
collaboration with Gramene	Gene trees	EPIGT00940000164429 (	Plant Compara)			
	TraesCS3D01G273	600				
	Description	Ensembl Genomes gene	TraesCS3D01G273600 is no longer in	the database. It has been mapped to 1 current identifier	(eg TraesCS3D02G273600).	
	Gene ID	TraesCS3D01G273600				
	Species	Triticum aestivum				
	Ensembl Plants release 42	- Dec 2018 © <u>EMBL-EBI</u>				
	About Us		Get help	Our sister sites	Follow us	
	About us		Using this website	Ensembl	Blog	
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	Disclaimer					

## Figure 4: Search Results

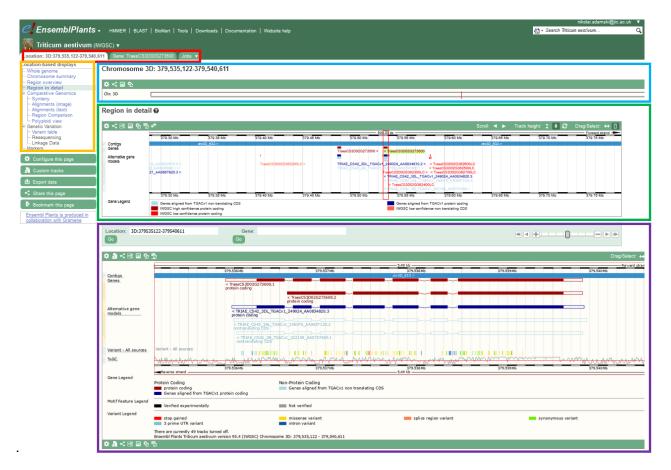
Querying the database opens a window with search results. From every result, you can navigate to a different part of the database.

If we follow the "Location" link, we are directed to the second layer of the Ensembl Plants website: The Location tab (red box in **Figure 5**). This is a genomic view of the region matching with the users query. The tab consists of three windows representing three different "zoom" levels:

The first window displays the genomic location relative to the entire chromosome (blue box in **Figure 5**). It allows the user to quickly browse between different positions on the chromosome and also to look at wider or narrower stretches of the chromosome. In case of the region being a scaffold (scaffolds are not part of the chromosome pseudomolecules!) this window only shows the scaffold itself.

The second window (by default) represents a region of 500 kb surrounding the users query. It displays the genes present within the 500 kb interval and thus allows the user to see the gens that for example underlie a quantitative trait locus (QTL). Other tracks, e.g. alternative splice variants, can be added as well.

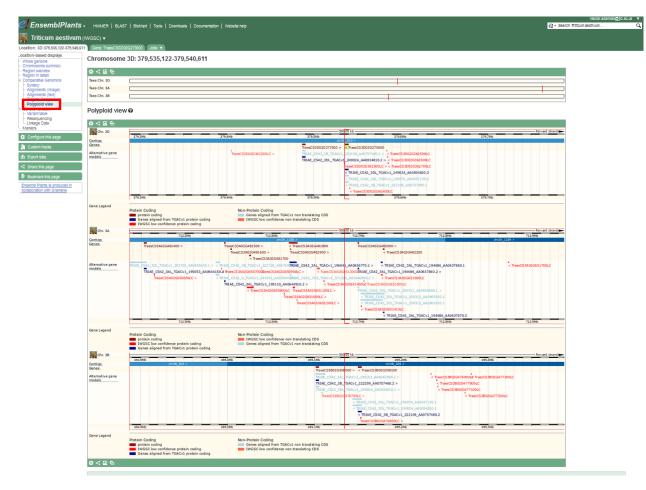
Finally, the third window represents a detailed view of the user's query, e.g. a gene model. By default, only the gene model and a track for sequence variation are shown here, but this window has a lot of extra tracks to offer (many more than the previous two windows). Users are encouraged to play with the available tracks as there is a lot of information available that is hidden by default.



## Figure 5: The Location tab

This tab displays the genomic region matching the users query. It is subdivided into three windows, each with a different "zoom" level. The pane on the left offers alternative views of the genomic region.

In addition to the three windows, there is also a pane on the left-hand side of the tab that allows users to view the region differently (orange box in **Figure 5**). In general, this pane offers tab specific viewing options, as we will see later on. One of these alternative viewing options is the "Polyploid view", which adds windows for the homoeologous regions, i.e. instead of the three windows described above a set of nine windows is displayed. This can be quite useful as it allows scanning of the three homoeologous regions quickly on different levels of detail.



## Figure 6: Polyploid view

When selecting "Polyploid view" from the left-hand side menu of the Location tab (red box in Figure 6) users can inspect genomic regions from all three homoeologs at the same time.

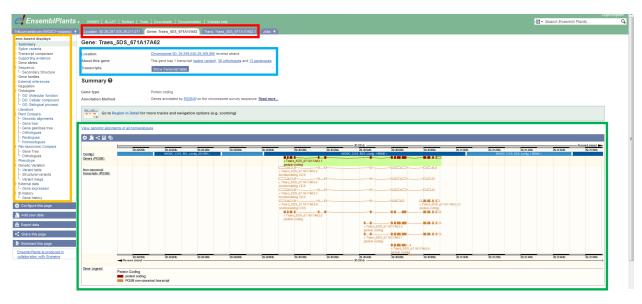
## The Gene tab

Going back to the query results in **Figure 4** users can look directly at a gene that falls within their query as opposed to the genomic location. For this, the user simply follows the "Gene ID" in the search results. Alternatively, one can get to the Gene tab after inspecting the genomic region by selecting the Gene tab in the bar above the information windows (see red box in **Figure 7**).

The Gene tab offers some basic information about the gene model in question, e.g. its genomic location or the number of splice variants (blue box in **Figure 7**). A single window (green box in **Figure 7**) shows detailed information on the gene and its surrounding genomic location. This window is similar to the detailed view window in the Location tab (purple window in **Figure 5**). The difference is mostly in the type of additional tracks that can be activated. Again, by default only the main gene model is being shown, but additional tracks like transcriptome data or alternative splice variants are also available.

As in the Location tab there is a pane on the left-hand side of the screen with various display options. By comparing **Figure 5** with **Figure 7** one can immediately see that there are many more display options in the pane of the Gene tab. As was mentioned before, these display options are tab specific. The available options depend on the data deposited on Ensembl Plants and contain for example simple GO terms of the gene or a detailed gene tree with homoeologous and

orthologous genes or a table listing the recorded variants of this gene (see section "<u>Variation data</u>" for more details).



## Figure 7: The Gene tab

This screenshot from the Ensembl Plants website shows the detailed gene view tab. This window displays some information about the gene model, e.g. its genomic location or the number of splice variants (blue box). The main window shows detailed information about the gene and additional tracks can be added (green box). The pane on the left side of the tab offers additional information, e.g. the names of homoeologous genes or GO terms (orange box). Note that the options in this pane are different to the ones in the Location tab. Users can navigate between e.g. the location and gene view using the tabs above the information windows (red box).

## The Transcript tab

The last of the tabs gives detailed information on the transcript of a gene model. Again, some basic information like the genomic location and number of splice variants is displayed as well as a very detailed view of the intron-exon structure without any genomic information (blue box in **Figure 8**). The strength of this tab is again in the different viewing options offered by the pane on the left-hand side of the screen (orange box in **Figure 8**). This offers for example a detailed cDNA view of the transcript, a table with variants or a list of the known protein domains encoded by the transcript.

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Triticum aestivum (WGSC1+popseq)	) ▼ Location: 5D:29,297,035-29,311,	071 Gene: Traes_5DS_671A17A62 Trans: Traes_5DS_671A17A62.1 Jobs 🔻								
ranscript-based displays - Summary	Transcript: Traes_5DS_671A17A62.1									
<ul> <li>Supporting evidence</li> <li>Sequence</li> </ul>	Location	Chromosome 5D: 29,299,340-29,309,066 reverse strand.	Chromosome 5D: 29.399.340-29.399.066 reverse strand.							
- Exons	About this transcript	This transcript has 15 axons, is annotated with 16 domains and features and is associated with 10 variations.								
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- Variant table - Variant image										
Population comparison Comparison image										
- Protein summary - Domains & features - Variants - ID History	Statistics Type	Exons: 15 Coding exons: 15 Transcript length; 2,124 kps Translation length; 612 residues								
- Transcript history	Annotation Method	Genes annotated by PGSB # <sup>2</sup> on the chromosome survey sequence. Read more	Novel protein coding							
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Ensembl Plants is produced in collaboration with Gramene	Citing Ensembl Genomes	Adding custom tracks	Ensembl Fungi							
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	EMBL-EBI Terms of use									

## Figure 8: The Transcript tab

This screenshot from the Ensembl Plants website shows the detailed transcript view tab. The tab consists of a window displaying information about the transcript, e.g. the UniProt ID of the translated protein or the annotation method as well as a view of the exon-intron structure (blue box). The pane on the left-hand side offers different display options (orange box).

# c) The power of Compara

Comparative studies are a powerful approach to identify genes in related species with specific functions, e.g. flowering time, plant height or grain size. These genes have likely been characterised in a model organism like rice and can be used as candidates to identify corresponding pathways in related but less well characterised species like wheat.

To identify the correct orthologous genes between species can be a daunting task depending on the size and similarity of the gene family as well as the number of genes a researcher is interested in. Luckily, the Ensembl Plants team has already done this work. As mentioned in <u>section b</u> the gene models on the Ensembl Plants website can be viewed in a gene tree format. These gene trees were built using a sophisticated pipeline (for more information on building the gene trees see <u>http://plants.ensembl.org/info/genome/compara/homology\_method.html</u>). Two types of trees were built where possible:

The Plant Compara tree compares genes from mostly the plant kingdom, although genes from other eukaryotic organisms are also included. The Plant Compara tree contains many more species than the Pan-taxonomic tree and is thus the tree of choice for plant to plant comparisons.

The Pan-taxonomic Compara trees are similar to the Plant Compara trees but include taxonomically more diverse species. Thus, this tree is better suited for comparing genes across kingdoms.

To access the gene trees users can either query the database with a gene name (see **Figure 9**) or select the gene tree view from the Gene tab display options panel (see orange box in **Figure 7**).

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Export data	Gene ID	<u>OS05G0113900</u>		
	Species	<u>Oryza sativa Japonica</u>		
Share this page	Location	<u>5:738208-739079</u>		
🛃 Bookmark this page	Gene trees	EPIGT00820000103317 (Plant Compara) EGGT00310000210695 (Pan-taxonomic Compara)		
Ensembl Plants is produced in collaboration with Gramene				
	Ensembl Plants release	≥ 31 - March 2016 © <u>EBI</u>		

#### Figure 9: Query results for a gene

This screenshot shows the query results for a gene from rice. From here, the user can either navigate to the Plant Compara tree or a Pan-taxonomic tree.

## The Gene tree view

The gene tree view window is separated into two major units. The first is a summary of the gene tree, e.g. how many species and genes are included (blue box in **Figure 10**) and how many of these genes share certain GO and INTERPRO domains (green box in **Figure 10**); the latter can be used to highlight genes with specific GO and INTERPRO domains.

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a sativa Japonica (IRGSP-1.0)	<ul> <li>Location: 5:738,208-739,07</li> <li>Gene</li> </ul>	: Os05g0113900 Trans: OS05T0	13900-01 Jobs 🔻		
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ne alleles	Location	Chromosome 5: 738,208-739,0	19 forward strand.		
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Secondary Structure ne families	Transcripts	Show transcript table			
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erature Int Compara		405			
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Sene tree	Number of duplication nodes	333			
Bene gain/loss tree Orthologues	Number of ambiguous nodes	12			
Paralogues	Number of gene split events	0			
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## Figure 10: Gene tree view part 1

This is the first screenshot of the gene tree view. Note that it is part of the Gene tab (red box). The first half of the tree view contains information on the gene tree (blue box) as well as on the GO and INTERPRO domains associated with the tree member (purple box).

The second part of the gene tree view consists of the actual gene tree and the alignments that were created using the Compara pipeline, with the initially queried gene being highlighted in red. By default, most of the gene tree is collapsed; users can then expand the sections they are Ensembl Plants primer 10
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interested in. Right-clicking a gene name opens up a tool tip giving user more information about the gene and also allowing users to follow a link to the gene. Both Plant Compara and Pantaxonomic gene trees share the same layout.

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-	Nesangiospermae: 115 homologs			
Pentapetala	e: 2 homologs			
	Musa acuminata: 8 hemologs			
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	<ul> <li>BRADI 1g30337v3, Brachypodium</li> </ul>			
	ONIVA06G26190, Oryza niva			
	CRUFID6G23930, Oryza rufip			
	BGIOSGA020813, Oryza sativ			
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	OMERI06G22190, Oryza meridior			
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	<ul> <li>OB06G29960, Onyza brachyanths</li> </ul>			
	Panicoideae: 3 homologs			
Hel	ianthus annuus: 2 homologs			
	Flowering plants: 91 homologs			
	PHAVU 008G185400g, Phaseolus vulgans			
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- x100 branch length		<ul> <li>duplication node</li> </ul>	<ul> <li>collapsed (gene of interest)</li> </ul>	■ 33 - 60% aligned AA ■ 66 - 100% aligned AA
		<ul> <li>ambiguous node</li> <li>gene split event</li> </ul>	(gene of interest)	<ul> <li>OV - 1007# dilgred 701</li> </ul>

## Figure 11: Gene tree view part 2

This screenshot shows the second part of the gene tree view. This contains the actual alignments on the right-hand side and the gene tree on the left. The initially queried gene is highlighted in red. By default, most of the tree is displayed in a collapsed fashion; users can expand the parts of the tree that is relevant to them.

Using these gene trees allows researchers to quickly identify orthologous genes in other species. In **Figure 10** and **Figure 11** we used a gene from rice as query and using the gene tree we can rapidly identify the orthologous genes from wheat.

The accuracy of the gene trees however depends on the quality of the gene models used. Although the current set of gene models for wheat (IWGSC RefSeqv1.1) are of very high quality, there is no guarantee that every gene is correctly annotated.

As a result, a wrong gene model can lead to an inaccurate gene tree. The example above highlights this nicely, as orthologous genes in wheat are apparently only found on chromosome 7B and 7D, but not 7A. While it is possible that the A-genome has lost its copy of this gene it is also quite possible that the 7A gene model is missing. A manual BLAST search using the genomic sequence of either the B- or D-genome gene model confirms that indeed the A-genome gene copy exists but is missing from the RefSeqv1.1 gene model set. Users are encouraged to check the wheat gene models using the criteria outlined in the "Gene models" section.

# d) Using BLAST on Ensembl Plants

As was mentioned at the start of the document users can query the Ensembl Plants database using BLAST. The BLAST tool here allows up to 30 sequences (either DNA or protein; blue box in **Figure 12**) to be used as queries against databases of up to 20 different plant species (green box in **Figure 12**). For every sequence and species, a separate job is automatically submitted, and users can view them later one by one. The BLAST parameters can be changed to suit a user's needs, e.g. e-value threshold, gap open penalties, etc (purple box in **Figure 12**).

C. EnsemblPlants	- HMMER   BLAST   BioMart   Tools   Dow	
Triticum aestivum (IWGSC1+popseq)	BLAST/BLAT	
Web Tools	BLAST search 🛛	
Variant Effect Predictor     Assembly Converter     ID History Converter	Sequence data:	Maximum of 30 sequences (type in plain text or FASTA)
<ul> <li>Configure this page</li> <li>Add your data</li> </ul>		
Add your data		Or upload sequence file Browse No file selected.
Share this page		ONA
🖈 Bookmark this page		O Protein
Ensembl Plants is produced in collaboration with Gramene		
	Search against:	Triticum aestivum
		Add/remove species
		DNA database     Genomic sequence
		Protein database     Proteins
	Search tool:	BLASTN
	Search Sensitivity:	Normal
	Description (optional):	
	General options	
	Scoring options	
	Filters and masking options 🖲	
		Run > Clear

## Figure 12: BLAST tool on Ensembl Plants

This is a screenshot from the Ensembl Plants BLAST tool. Up to 30 sequences (blue box) can be used as queries against up to 20 species (green box). The BLAST parameters can be tweaked to achieve best results (purple box).

The output of a BLAST search is displayed in a table with columns for alignment length, percent similarity, etc (see **Figure 13**). The table can be downloaded (like all tables on Ensembl Plants) and the pair-wise alignments can be accessed by clicking the "Alignment" link in the percent identity column.

Every HSP (High-scoring Segment Pair) is reported as a separate hit, so using CDS (coding sequence) as query to BLAST a genomic database will result in a lot of hits. Thus, it is often easier

to interpret the results as there will be fewer high-quality hits when using genomic sequence for BLAST queries on Ensembl Plants.

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m aestivum (IWGSC1+popseq) V BLAST/BLAT V							
Tools Results for Vrn1_gen	omic @						
LAST							
Tidet Job details							
rient Effect Predictor Job name	Vrn1_genomic						
embly Converter fistory Converter	/=						
igure this page	Triticum aestivum						
Assembly	IWGSC1+popseq						
	BLASTN (NCBI BLAST)						
t data							
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nork this page Results table							
of Plants is produced in retion with Gramono		Show/hide columns (4 hidden)					Filter
Genomic Location		Overlapping Gene(s)	Orientation	Length	Soore	* E-val	%ID
IWGSC CSS 5AL scaff 28053		Traes 5AL 13E2DEC48	Forward	3344 [Sequence]	3276	0.0	99.5 (Alignment)
IWGSC CSS 5DS scaff 27780		Traes 5DS 8668AC690	Forward	1809 (Sequence)	1793	0.0	99.8 (Alignment)
IWGSC CSS 5DS scaff 25510		Trees 5DS B05590809	Reverse	1210 Decience	1178	0.0	99.3 (Alignment
IWGSC CSS 5AL scall 28053	9.11509-12098 (Sequence)		Forward	590 (Securical)	562	0.0	98.8 (Alternant)
IWGSC CSS 6DS scall 25510			Reverse	590 (Sequence)	562	0.0	98.8 (Allowert
IWGSC CSS 5BL scott 10898	38 8530-9183 (tecueros)	Trees 5BL 502022E67	Reverse	664 (tecuence)	538	0.0	95.6 (Allowert)
5D-129721968-129722368 (Sec)	225	Trees 5DL 9CO4EC839	Forward	401 (sequence)	329	0.0	95.5 (Allowed)
IWGSC CSS 5AL scaff 28053	19:11008-11314 (Becuence)	Trees 5AL 13E2DEC48	Forward	307 (Sequence)	307	1.3E-170	100.0 (Alignment)
IWGSC CSS 5DS scaff 25510	15:2280-2585 (Sequence)		Reverse	307 (Sequence)	307	1.3E-170	100.0 (Alignment)
5D:129723075-129723448 [Sec.	noi	Traes 5DL 9CC4EC839	Forward	372 (Sequence)	304	7.9E-169	95.4 (Alignment)
IWGSC CSS 58L scaff 10898	38:7041-7412 (Sequence)	Traes 58L 502022E67	Reverse	372 (Sequence)	300	1.9E-168	95.2 (Alignment)
5D.129721056-129721398 [Sec.	nos)	Traes 5DL 9CC4EC839	Forward	343 (Securice)	287	1.1E-158	95.9 (Alignment)
5D.129721487-129721813 (Sec.	101	Trees 5DL 9CC4EC839	Forward	327 (Securical)	283	2.7E-158	98.6 (Alignment)
WGSC CSS 58L scall 10898	38.9500-9780 (Sequence)	Trees 5BL 502022E07	Reverse	281 (Securical)	249	5.2E-138	97.2 Migmint
5D:129720459-129720736 (Sec)	100	Trees 5DL 9CC4EC839	Forward	278 (Sequence)	242	7.9E-132	96.8 (Algoment)
5D:129720741-129721054 (Sec)	222	Trees 5DL 9CC4EC839	Forward	314 (tequence)	242	7.9E-132	94.3 (Algoment)
5D:129722787-129723071 (Sec)	100	Trees 5DL 9CC4EC839	Forward	285 (Sequence)	221	2.7E-119	94.4 (Algoment)
IWGSC CSS 5BL scaff 10898	38:7415-7700 (Sequence)	Traes 58L 502022E67	Reverse	286 (Sequence)	218	1.7E-117	94.1 (Alignment)
IWGSC CSS 58L scaff 10898	38.9185-9498 (Becuerce)	Trees 58L 502022667	Reverse	314 (Sequence)	214	4.0E-115	92.0 (Alignment)
						2.3E-107	
IWGSC CSS 5BL scall 10898	[38:8289-8529 [Sequence]	Traes 58L 502022E67	Reverse	241 (Sequence)	201	2.3E-107	95.9 (Alignment)

## Figure 13: BLAST results

This screenshot shows the output of a BLAST search on Ensembl Plants. The output is in tabular format that can be downloaded. Genes overlapping genomic sequences are shown in a separate column.

# e) Using BioMart on Ensembl Plants

At the beginning of this document we mentioned the tool BioMart. This is essentially a tool to download specific sets of data in both small and large quantities.

Users first select the type of data and organism they are interested in (see **Figure 14**). Then they can apply a set of filters, e.g. a genomic location, to limit the number of results retrieved from their query: Setting no filter would mean that all data would be downloaded. Furthermore, the user can specify what attributes the data should have, e.g. GO terms associated with genes. Lastly, a user can specify an additional dataset from a different organism, e.g. rice, for which filters and attributes can again be set separately; using two datasets allows users to download for example all wheat genes and their orthologous genes from rice. Thus, by combining different sorts of filters, attributes and datasets individual users can download data specifically tailored to their needs.

The number of filters and attributes that can be used is big and can be daunting for new users. It is highly recommended for first time users to download small sets of defined data to get used to the interface. Once users understand the basic concepts however BioMart offers quick access to large quantities of data.

We have created a step-by-step example of how to use BioMart, which can be found in the **Finding wheat orthologs** section.

<i>C</i> EnsemblPlants	
New Count Results	
Dataset	Plant Mart -
Triticum aestivum genes (IWGSC1+popseq (2.2))	Triticum aestivum genes (IWGSC1+popseq (2.2))
Filters	
[None selected]	
Attributes	
Gene stable ID	
Transcript stable ID	
Dataset	
[None Selected]	

**Figure 14: BioMart on Ensembl Plants** This screenshot shows the basic layout of the BioMart tool on Ensembl Plants. Users can select datasets from different species, select filters and choose what type of data they want to download.